

## DATA SHEET

### eFISH CEN X/Yq12 Dual Color Probe

#### Catalog No.

**FP102-10X-100µl- 10 test**

**FP102-20X -200µl- 20 test**

Doc No: 932-FP102 Rev: C

Date of Release: 11-Jul-2024

Material Provided: One vial of eFISH probe in hybridization buffer (RTU).

#### Recommended detection system (Not supplied):

Either of the following detection systems is recommended depending on the automation/manual platform used:

eFISH Kit	Cat #	Description
eFISH Histo	DF-500-20X	Automation
eFISH Cyto	DF-510-20X	Automation

#### Intended Use:

The BioGenex eFISH CEN X/Yq12 Dual Color Probe is currently available for Research use only. eFISH eFISH CEN X/Yq12 Dual Color Probe is designed to be used for the detection of human chromosome X alpha-satellites as well as chromosome Yq12 specific sequences in formalin-fixed, paraffin-embedded tissue or cells by fluorescence in situ hybridization (FISH).

eFISH BioGenex CEN X/Yq12 Dual Color Probe in hybridization buffer. The probe contains orange labeled polynucleotides (Orange: excitation at 547 nm and emission at 572 nm, similar to rhodamine), which target alpha-satellite sequences (DXZ1) in the chromosomal region Xp11.1-q11.1, and green-labeled polynucleotides (Green: excitation at 503 nm and emission at 528 nm, similar to FITC), which target sequences of the satellite III region (DYZ1) of chromosome Y in the chromosomal region Yq12.

#### Summary and Explanation

Fluorescence *in situ* hybridization (FISH) is a robust technique of cytogenetics used for the detection of chromosomal aberrations, presence or absence of specific DNA sequence in native context. In this technique fluorescent probes bind to the target sequence of DNA in chromosome. High specificity and sensitivity coupled rapid and an accurate result has proven role of FISH in both research and diagnosis of solid tumor and hematological malignancies. As technique of cancer cytogenetics, FISH, can be used to identify genetic aberrations viz., deletions, amplification and translocation in tissue sections or within

individual cells. FISH is also used for use in genetic counseling, medicine, and species identification. FISH can also be used to detect and localize specific RNA targets in cells, circulating tumor cells, and tissue samples<sup>1,2,3,4,5</sup>.

In FISH procedure, fixed tissue sections are pretreated to expose target DNA or mRNA sequences. An appropriately labeled probe is hybridized to the exposed target DNA or mRNA sequences in the cells. Subsequent stringent washing steps remove any probe that is non-specifically bound to the tissue section. Subsequently slides are mounted using DAPI/antifade and can be visualized under fluorescence microscope using appropriate filter set.

#### Principles of the Procedure

*In Situ* hybridization (ISH) allows the detection and localization of definitive nucleic acid sequences directly within a cell or tissue. High specificity is ensured through the action of annealing of fluorescence probe nucleic acid sequence to complementary target nucleic acid sequence. ISH techniques can be used to identify genetic aberrations like deletions, amplification, and translocation in tissue sections or within individual cells.

#### Storage and Handling

The BioGenex eFISH CEN X/Yq12 Dual Color Probe must be stored at 2-8°C protected from light and is stable through the expiry date printed on the label.

#### Specimen Collection and Slide Preparation

Tissues fixed in 10% (v/v) formalin are suitable for use prior to paraffin embedding and sectioning.

#### FISH Staining procedure

- The BioGenex eFISH probes are supplied in hybridization buffer and used without further dilution.
- Protocol:

Please refer to the eFISH probe specific instruction/protocol for automated or semi-automated FISH processing platform (Xmatrx<sup>®</sup>-Infinity, Xmatrx<sup>®</sup>-Nano and Xmatrx<sup>®</sup>-mini).

Further processing, such as washing and counter-staining, can be completed according to the user's needs. For a particularly user-friendly performance, we recommend the use of a BioGenex eFISH kit.

**Disclaimer:** The above information is provided for reference only. Each end-user is responsible for developing and validating optimal testing conditions for use with this product.

### **Troubleshooting**

Contact BioGenex Technical Service Department at **1-800-421-4149** or your local **distributor** to report unusual staining.

### **Expected Results**

For BioGenex eFISH CEN X/Yq12 Dual Color Probe, the hybridization signals of the labeled alpha-satellite-sequences of the centromere of chromosome X appear orange; the hybridization signals of the labeled chromosome Yq12 specific sequences appear green.

In interphases of normal male cells or male cells without aberrations of chromosome X and Y, one chromosome X and one chromosome Y signal appears. In interphases of normal female cells or female cells without aberrations of chromosome X, two chromosome X signals appear. In cells with aneuploidy of chromosome X or Y, a different signal pattern is visible in interphases.

Care should be taken not to evaluate overlapping cells, in order to avoid false results, e.g. an amplification of genes. Due to decondensed chromatin, single FISH signals can appear as small signal clusters. Thus, two or three signals of the same size, separated by a distance equal to or less than the diameter of one signal, should be counted as one signal.

### **Limitations of the Procedure**

Correct treatment of tissues prior to and during fixation, embedding, and sectioning is important for obtaining optimal results. Inconsistent results may be due to variations in tissue processing, as well as inherent variations in tissue. The results from *in situ* hybridization must be correlated with other laboratory findings.

### **Bibliography**

1. Gall, J. G. and Pardue, M. L. (1969). *Proc. Natl. Acad. Sci. USA* 63, 378 -383.
2. Rudkin, G. T. and Stollar, B. D. (1977). *Nature* 265, 472-473.
3. Hougaard, D. M., Hansen, H. and Larsson, L. I. (1997). *Histochem. Cell Biol.* 108, 335 -344.
4. Bauman, J. G., Wiegant, J., Borst, P. and van Duijn, P. (1980). *Exp. Cell Res.* 128, 485 -490.
5. O'Connor, C. (2008). *Nature Education* 1(1):171.